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<b>(54) Title:</b> ENHANCEMENT OF IMMUNOCHEMICAL STAINING IN ALDEHYDE-FIXED TISSUES  <b>(57) Abstract</b>  A method for restoring immunoreactivity of a tissue, particularly decalcified tissue, fixed with an aldehyde fixative agent and embedded in an embedding medium, usually comprising celloidin, the method comprising the steps of contacting the tissue with an aldehyde releasing reagent solution comprising a solvent and an aldehyde releasing reagent and removing aldehyde released by the aldehyde releasing reagent from contact with the tissue by reacting the aldehyde in a substantially irreversible manner to form a non-aldehyde derivative, and removing excess base from the tissue. A preferred solution for celloidin-embedded decalcified tissue comprises methanolic sodium hydroxide at about one-third saturation.		

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## ENHANCEMENT OF IMMUNOCHEMICAL STAINING IN ALDEHYDE-FIXED TISSUES

The invention concerns immunohistochemical staining of aldehyde-fixed and embedded tissue sections. Tissue sections obtained from clinical specimens or animal experimentation frequently are fixed, embedded, and stored in a form suitable for later examination by light microscopy. Immunological reagents, especially monoclonal antibody reagents, currently permit examination of at least certain of these fixed tissue samples for the presence of particular antigenic compounds. Antigens of interest may be associated with a disease process or pathology, or may identify a particular cell type or tissue. In the case of recently prepared biopsy and autopsy samples, such immunohistochemical analyses are of immediate diagnostic value.

However, immunohistochemical analyses of tissue specimens have been hampered because of antigenic loss during specimen fixation. Traditional fixation methods frequently have employed aldehyde fixatives, which fix the tissue by causing cross-linking reactions within and between tissue proteins.

Two types of cross-linking reactions have been recognized. The first is a Schiff's base-type polymerization: formaldehyde condenses with the amino groups of the protein, resulting in the Schiff's base intermediate, which is capable of undergoing rapid polymerization leading to cross-linking of the proteins.

In the second type of reaction, called the Mannich reaction, the formaldehyde can react with both an amino group and an active hydrogen group, resulting in the formation of a Mannich base. Polymerization of the Mannich bases results in protein cross-linking.

Cross-links preserve tissue morphology and integrity, harden the tissue for slicing, and inhibit microbial attack. Unfortunately, the cross-linking process also causes loss of tissue antigenicity, a result which impedes the usefulness of immunological reagents on tissues fixed with aldehyde reagents such as formaldehyde. The chemistry of the cross-linking of amino acids and proteins by formaldehyde is described in Harlan and Fearheller, "Chemistry of the Cross-

Linking of Collagen During Tanning," and Kelly, et al. "Cross-Linking of Amino Acids By Formaldehyde," (1976). The role of Mannich-type reactions in cross-linking of protein amino groups and aromatic amino acids with formaldehyde is discussed in Fraenkel-Conrat, et al., J. Biol. Chem. (1947) 168:99-118, and  
5 Fraenkel-Conrat and Olcott, J. Biol. Chem. (1948) 174:827-843. Further discussions of aldehyde cross-linking reactions are found in Fox, J. Histochem. Cytochem. (1985) 33:845-855; Jones, "Reactions of aldehyde with unsaturated fatty acids during histological fixation," in Fixation in Histochemistry, P. J. Stoward, ed. (1973); and Kunkel et al., Mol. Cell. Biochem. (1981) 34:3.  
10 Mannich type reactions are described in general in March, "Advanced Organic Chemistry," particularly at 333, 424, 670-672 (1968).

In an attempt to circumvent the disadvantages of aldehyde fixation, alternative fixation methods have been developed, such as microwave heating (Mayers, J. Clin. Pathol. (1970) 28:273; Hopwood et al., Histochem. J. (1984)  
15 16:1171) and alcohol immersion (Battifora and Kopinski, J. Histochem. Cytochem. (1986) 34:1095). Despite some advantages of alternative fixation methods, they have not displaced aldehyde fixation in general use. Their limited acceptance may reflect drawbacks present in these alternative methods. For example, microwave heating lyses red cells and disrupts membrane lipids.  
20 Although ethanol fixation is reported to produce improved antigenicity of tissue samples, ethanol causes increased cellular shrinkage (Battifora and Kopinski, id.) Consequently methods for restoring antigenicity to aldehyde fixed tissues continue to be useful for specimens generated by current clinical practices.

In addition, methods for restoring antigenicity are useful because of the  
25 vast number of aldehyde fixed tissue samples already in collections. These stored tissue samples provide a rich reservoir of material for retrospective immunohistochemical examination. If a suitable method of subsequent immunohistochemical staining were available, newly generated immunohistochemical data could be combined with existing diagnostic results obtained from traditional  
30 investigations on the same tissues. Often clinical samples are saved for decades, so that the clinical outcome of the patient's underlying pathological process already is known. In the case of experimental tissues, such as those obtained from

animals in toxicology testing, other measurements of pathology and toxicity in general already will have been performed and documented. In both cases, immunological analyses of the affected tissues could add important correlative information.

5       Because of the development of immunological reagents over the past decades, immunohistochemical analyses can now be performed that were impossible at the time many tissues were originally stored. In addition, new knowledge or hypotheses concerning the disease process may prompt reexamination of stored tissues. Immunohistochemical studies on stored tissue  
10       samples provide a relatively time- and cost-effective means for performing a clinical study on a statistically large sample population. Therefore the application of immunohistochemical analyses to routine clinically or experimentally derived embedded tissue sections is a matter of considerable interest.

      Antigenic loss during aldehyde tissue fixation is due to chemical  
15       modification of the protein (not to physical removal of the antigen). Loss of immunoreactivity is believed to occur by two mechanisms. In the first mechanism, the fixative agent chemically modifies the reactive epitope, rendering it incapable of binding antibody. In the second mechanism, the fixative agent causes chemical cross-linking of the antigenic protein at sites outside the targeted  
20       epitope. Such cross-linking may be intramolecular or intermolecular, i.e., with involvement of nearby proteins. This cross-linking sterically hinders access of the antibody reagent to the reactive epitope.

      The second mechanism, steric hindrance due to intraprotein or interprotein cross-linking outside the epitope of interest, has been reversed by protease  
25       digestion of formalin fixed tissues in order to remove the interfering cross-linked portions of proteins. This approach has been shown to improve immunostaining of keratins in formaldehyde fixed tissues (Battifora and Kopinski, J. Histochem. Cytochem. (1986) 34:1095-1100). However, protease treatment actually degraded the staining of tissues fixed in alcohol, a fixative solution which does not  
30       cause protein cross-linking. Some improvement in immunostaining by proteolysis of formaldehyde fixed tissues for limited time periods has been shown for other antigens (Huang, et al., J. Lab Invest. (1976) 35:383-390.) In a study of

immunostaining of 23 antigens of pathological interest, prior trypsinization was shown to give no improvement in immunostaining of formaldehyde fixed tissues except in the case of cytokeratins and desmin. For many antigens, enzyme digestion actually diminished antigen staining. (Leong, et al., J. Pathology (1988)

5     156:275-282.)     These results support the hypothesis that aldehyde fixative-induced cross-linking of proteins diminishes immunostaining both by chemical modification of epitopes and by steric hindrance mediated by cross-linking. The mixed results obtained from treating fixed tissues with proteolytic enzymes are readily rationalized: although partial proteolysis decreases cross-linking and  
10     reduces steric hindrance, proteolysis also may cleave and remove the epitopes of interest. Moreover, proteolysis cannot reverse antigenic masking due to chemical modification of the epitope. By contrast, a procedure which could reverse the chemical cross-linking reaction produced by aldehyde fixatives has the potential to unmask antigens previously hidden by either mechanism.

15     A procedure for restoring antigenicity of formalin-fixed, paraffin embedded tissue sections by heating the tissue in a microwave in a heavy metal solution has been described in Shi, et al. J. Histochemistry and Cytochemistry 39(b): 741-48 (1991). This procedure provides enhanced immunostaining in approximately three-fourths of the samples tested. The described method is part of a process that  
20     involves the steps of tissue section deparaffinization and rehydration, brief treatment with aqueous peroxide to block endogenous peroxidase, washing of the slides with distilled water, covering the slides with distilled water or a heavy metal solution, and brief microwave heating for several minutes. Following this procedure, slides are cooled, rinsed, and immunostained in a conventional fashion.

25     This method for restoring antigenicity is subject to certain limitations. First, it requires the use of a microwave oven to heat the tissue samples. Many laboratories may not be equipped with a microwave oven, and some tissue samples may not be suited to microwave heating. A need exists for an antigen retrieval  
30     method that can be used at room temperature, without any external heat source. In addition, the previously described procedure is especially suitable for tissues embedded in a hydrocarbon medium such as paraffin. It is not well suited for

tissue sections embedded in celloidin, a preferred embedding medium for bony tissues. A need also exists for a method which is suitable for use with celloidin embedded tissues. Moreover, a need particularly exists for a method which may be used with decalcified bony tissue samples, since decalcified tissues are often refractory to the previously described method.

Methods and compositions are provided for restoring antigenicity for immunohistochemical analysis of aldehyde fixed tissue. The methods and compositions are especially useful with celloidin embedded tissue and with decalcified bony tissue. The method involves treating aldehyde fixed tissue with a chemical agent or agents that catalyze an essentially irreversible Mannich/Schiff's Base reaction which converts the aldehyde released thereby into a nonreactive form, i.e., into a non-aldehyde derivative. In this application such chemical agent or agents will be termed an "aldehyde releasing agent". In one aspect, the aldehyde releasing reagent comprises a nucleophilic reagent solution and the method involves (1) treating aldehyde fixed tissue with the nucleophilic reagent solution, optionally containing a chaotropic agent, (2) removing excess nucleophile by neutralizing or rinsing the tissue sample, and (3) reacting the tissue with an immunoreactive reagent. In another aspect, the aldehyde releasing reagent comprises an oxidizing agent and the method involves (1) treating aldehyde fixed tissue with the oxidizing agent, (2) removing excess agent and (3) reacting the tissue with an immunoreactive reagent. In a third aspect, the aldehyde releasing reagent comprises an organic acid/base pair and the method in step (1) involves treatment with the organic acid/base pair. Exemplary organic acids are set forth below. Exemplary bases are sodium hydroxide and potassium hydroxide. Also provided is a kit for immunostaining of aldehyde fixed tissue. The kit minimally comprises a solution for restoring antigenicity comprising a solvent and an aldehyde releasing reagent. Optionally, the kit can also comprise a solution to remove excess aldehyde releasing reagent or a reagent immunostaining reagent.

The invention provides a method for restoring immunoreactivity of a tissue fixed with an aldehyde fixative agent and embedded in an embedding medium, the method comprising the initial step of contacting the tissue with a solution for restoring antigenicity comprising a solvent and an aldehyde releasing reagent. The

Most effective aldehyde releasing reagents for the practice of the invention are nucleophiles, preferably basic nucleophiles. An especially preferred nucleophile is hydroxide anion, which is conveniently supplied as an alkali metal hydroxide such as sodium or potassium hydroxide. Other convenient nucleophiles include primary, secondary, or tertiary amines, especially those with minimal steric hindrance to attack, such as piperidine or morpholine. Hydroxylamine and glycine are preferred. Other nucleophiles include thiols such as mercaptoethanol. Yet another nucleophile of interest is azide, e.g. sodium azide ( $\text{NaN}_3$ ). In general, any nucleophile capable of promoting a reverse Mannich reaction will be capable of cleaving at least some protein cross-linkages, as such reagents will also catalyze reversal of other types of reactions caused by formaldehyde. The concentration of the nucleophile may vary widely, with more concentrated solutions acting more quickly. For short exposures, nucleophile concentrations of 0.5M or greater are usually preferred. In the case of NaOH in methanol, concentrations of one-tenth to one-half of saturation (approximately 0.6 to 3M) are preferred in most circumstances. For hydroxylamine, for example in the form of hydroxylamine hydrochloride, or for glycine, a 10% aqueous solution is preferred.

Other effective aldehyde releasing reagents for the practice of the invention are oxidizing agents. Preferred oxidizing agents are hypochlorites and periodates, especially sodium hypochlorite or sodium periodate. Additional oxidizing agents include hydrazine hydrate. Concentration of the oxidizing agent will vary with the reagent. For example, for sodium hypochlorite, an aqueous (v/v) solution in the range of about 0.01% to about 0.005%, preferably 0.005%, is suitable. For sodium periodate, an aqueous (v/v) solution in the range of about 0.1% to about 1.0%, preferably 0.1%, is suitable. And for hydrazine hydrate, an aqueous (v/v) solution in the range of about 2.0% to about 5.0%, preferably 5.0%, is suitable. Treating the sample with the oxidizing agent is believed to break the cross-linkages between the aldehyde and tissue components and converts the released aldehyde into a non-reactive form, for example by converting formaldehyde to formic acid.

We have found that certain acid/base pairs will function as an aldehyde releasing agent within the confines of this invention. The following pairs are exemplary:



	<u>Organic Acid/Base Pair</u> <u>(Acid/Base)</u>	<u>Concentration</u> <u>(Acid/Base)</u>	<u>Minutes</u>
5	Trichloroacetic Acid/NaOH	10% aqueous/10% NaOH in methanol	10/20
	Tolvene Sulfonic Acid/NaOH	5% aqueous / 40% aqueous	30/30
	Citric Acid/NaOH	10% aqueous / 40% aqueous	30/30
	Oxalic Acid/NaOH	20% aqueous / 40% aqueous	30/30
10	Tartaric Acid/NaOH	10% aqueous / 40% aqueous	30/30

The solvent for the aldehyde releasing reagent solution may be any solvent compatible with and capable of dissolving the aldehyde releasing reagent.

Aqueous solutions are possible; and preferable where the aldehyde releasing reagent is an oxidizing agent or an organic acid/base pair. Organic solutions are preferable where the aldehyde releasing reagent is a nucleophile because they promote better penetration of the embedding medium. In addition, proteolytic fragments are insoluble in most organic solvents and therefore tend to remain in place on the slide.

A preferred solvent for use with celloidin embedded tissue sections is a polar organic solvent. Alcoholic solutions are preferred because they promote good penetration of the celloidin embedding medium and are good solvents for nucleophiles of interest, especially alkaline metal hydroxides. Lower alcohols are preferred, such as methanol, ethanol, propanol, and butanol; methanol is especially preferred. Polyols such as ethylene glycol and glycerol are also useful; they have the advantages of low volatility and greater viscosity, which permit them to remain on the slide for an extended period without evaporating or running off. Polar aprotic solvents such as dimethyl formamide (DMF) and dimethyl sulfoxide (DMSO) also may be used with appropriate nucleophiles. Furthermore, mixed solvent solutions are acceptable, provided the component solvents are compatible with the reagents and each other.

Two solvents may be used consecutively under some circumstances. For example, a celloidin embedded slide may be treated initially with methanol to

useful with celloidin, which is a traditional embedding medium for bony tissues. Celloidin is a pure form of pyroxylin, the low-nitrogen form of nitrocellulose. Celloidin is available from various commercial sources.

5 The described method preferably is performed with solution which either solubilizes or swells and softens the embedding medium. Most embedding media are supplied as solutions. Hence an appropriate solvent for the solution may be inferred based upon the solvation characteristics (e.g. hydrophobicity, polarity, hydrogen bond donor/acceptor potential) of the solvent supplied for the embedding medium. The solution need not employ a solvent identical to that used for the embedding medium. However, the identity of suitable embedding medium solvents provides a guideline to the appropriate characteristics for the aldehyde releasing solvent.

10 In the case of celloidin, the embedding medium is soluble in ether-alcohol mixtures, clove oil (comprising aromatic terpenes), alcohols, and acetone. Methanol is a suitable solvent, since it also solubilizes basic nucleophiles, and is a preferred solvent for use with celloidin embedded tissues.

Bony tissues in the past have presented particular problems, at least in part because of the decalcification treatment to which bone often is exposed during fixation. Conventional decalcification is performed with an acid such as trichloroacetic acid ( $\text{Cl}_3\text{CCOOH}$ ). At present, immunohistochemical staining is not widely used in diagnostic and investigative pathology for routinely processed formaldehyde fixed, decalcified, and celloidin embedded bony tissues, as for example temporal bone sections. Several reported attempts to immunostain temporal bone sections focus on modified fixation, decalcification, and embedding protocols (Veldman *et al.*, *Advances in oto-immunology*. New trends in functional pathology of the temporal bone, *Laryngoscope* (1987) 97:413; Huizing *et al.*, *Progress in temporal bone histopathology*. I. Semithin 3-5 um sectioning of undecalcified human temporal bone after plastic embedding, *Acta Otolaryngol (Stockh)* (1985) Suppl 423:24; Veldman *et al.*, *Progress in temporal bone histopathology*. II. Immuno-technology applied to the temporal bone, *Acta Otolaryngol (Stockh)* (1985) Suppl 423:29; Arnold, W., *Immunohistochemical investigation of the human inner ear*, *Acta Otolaryngol (Stockh)* (1988) 105:392;

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and Bauwens *et al.*, Progress in temporal bone histopathology. III. An improved technique for immunohistochemical investigation of the adult human inner ear, Acta Otolaryngol (Stockh) (1990) Suppl 470:34).

5 The disclosed method simultaneously neutralizes any residual acidity from decalcification, which might impede restoration of antigenicity using water or other solvent by itself with bony specimens. As the accompanying experimental results demonstrate, the disclosed method permits effective immunostaining of temporal bone sections which have been routinely processed and embedded. This is significant because human temporal bone collections amounting to 8,000 and  
10 13,000 specimens exist in Europe and the United States, respectively (Schuknecht, Ann. Otol. Rhinol. Laryngol. (1987) 96 (Suppl. 130):1). These collections provide an excellent research base for understanding otopathology by light microscopy.

A kit for use in immunostaining of a tissue can be provided to simplify practice of the method described above. The kit will minimally contain a  
15 receptacle adapted to hold one or more individual reagent containers and at least a first container containing (1) an aldehyde releasing reagent solution comprising a solvent and an aldehyde releasing reagent or (2) the aldehyde releasing reagent in an amount appropriate to make up the desired concentration when solvent from another container is used to fill the aldehyde releasing reagent container to a  
20 predetermined level. In most cases, the kit will also contain a second container containing (1) an immunostaining reagent or (2) a wash solution for removing excess aldehyde releasing reagent solution, or containers with both such materials. Wash solutions are typically buffered solutions that do not further dissolve or swell the embedding medium, such as aqueous buffered solutions. The solvent in the  
25 wash solution will be capable of dissolving the solvent and aldehyde releasing reagent used in the antigen retrieval solution. The immunostaining reagent generally comprises an antibody and staining moiety. Such reagents are well known in the art and require no further description here. Specific examples are given in the general examples of the invention set out below. Appropriate  
30 instructions for carrying out the method of the invention will also be included in the kit.

The invention now being generally described, the same will be better understood by reference to the following detailed examples which are provided for illustration and are not to be considered as limiting the invention unless so specified.

### EXAMPLE 1: A NUCLEOPHILE AS AN ALDEHYDE RELEASING REAGENT

#### 1. Materials and Methods

A total of 60 celloidin-embedded human temporal bone sections were obtained from the Eastern National Temporal Bone Bank at Massachusetts Eye and Ear Infirmary (Table 1). Most sections were processed routinely by either Heidenhain-Susa or 100% formalin fixation and decalcified by 5% trichloroacetic acid as described previously (Schuknecht HF. Pathology of the Ear. Cambridge, MA: Harvard University Press. 1974). Only one case was processed by a modified method using 10% neutral buffered formalin fixative and EDTA decalcification.

The monoclonal antibodies used are listed in Table 2. All antibodies were obtained from BioGenex Laboratories (San Ramon, CA). Most slides were stained with Super Sensitive biotin-streptavidin kits (SSBSA) from BioGenex. A few slides were stained by ABC kits purchased from Vector Laboratories, Inc. (Burlingame, CA).

#### 2. Preparation of Aldehyde Releasing Reagent Solution

Sodium hydroxide (NaOH) in methanol solution provides one formulation of the basic reagent used in these experiments. NaOH, 50-100 grams, was added to 500 ml methanol in a brown colored bottle. The solution was mixed by shaking and stored at room temperature for 1-2 weeks to settle. The upper layer of liquid was removed carefully from the precipitate, and diluted 1:3 in methanol. Prior to use, 0.01% sodium thiocyanate optionally was added.

#### 3. Treatment of Tissue Sections

The celloidin-embedded temporal bone sections were washed in distilled water for 10 minutes, mounted on 0.1% poly-L-lysine (Sigma) coated slides. The slides were immersed in one-quarter saturated NaOH-methanol solution (saturated methanolic NaOH, approximately 6M, diluted 1:3 with methanol) alone or with added 0.01% sodium thiocyanate, for 30 minutes. Slides were rinsed for 15 minutes in two changes of 100% and 70% methanol and two changes of phosphate buffer saline (PBS), followed by treatment with 0.3% Triton X-100 for 10 minutes and rinsing in PBS again.

#### 4. Immunostaining Procedures

Treatment was followed by a three-step immunostaining technique using either the SSBSA or the ABC method, as previously described (Shi et al., J. Histochem. Cytochem. (1991) 39:741). Briefly, slides were incubated with primary antibody overnight at room temperature followed by a 40 to 60 minute incubation with link (BioGenex Super Sensitive biotinylated anti-mouse immunoglobulin or Vector biotinylated anti-mouse immunoglobulin). Label (BioGenex Super Sensitive Alkaline phosphatase conjugated streptavidin or peroxidase conjugated streptavidin and Vector ABC) was added for 40 to 60 minutes. Slides were rinsed between incubations in three changes of PBS for 15 minutes. Either fast red or DAB chromogen was used as substrate. The immunostaining results were controlled by light microscopy.

The primary antibody was replaced with either nonspecific mouse ascites or PBS for negative control slides.

**Table 1.** Routine celloidin-embedded human temporal bone sections used  
(N = 60)

	Code	No. of sections	Fixation	Decalcification	Cut	Stored
5	P.F.	10 (L&R)	NBF	EDTA	Recent	< 1 yr
	C.B.	10(R)	H-S	TCA	1989	< 2yrs
	R.L.	10(L)	F	EDTA	1989	< 2yrs
10	G.H.	10(R)	H-S	TCA	1986	4 yrs
	J.P.	10(R)	H-S	TCA	1982	8 yrs
	R.Y.	10(L)	F	TCA	1960	30 yrs
	a.	L = left side. R = right side of temporal bone.				
15	b.	NBF = 10% neutral buffered formalin. H-S = Heidenhain-Susa fixative. F = 10% formalin.				
	c.	TCA = trichloroacetic acid.				

## 20 5. Results

The immunoreactivity of 15 monoclonal antibodies used on routinely processed celloidin-embedded sections is summarized in Table 2. The staining results showed strong positive staining for 7 monoclonal antibodies, moderate positive staining for four antibodies, and weak positive staining for one antibody. Three antibodies showed negative results. There was no significant difference in immunoreactivity between various sections. All negative control slides (PBS or nonspecific mouse ascites) showed negative staining. The intensity of immunostaining obtained by the SSBSA system was stronger than that obtained by the ABC system.

30 The immunoreactivity of monoclonal antibody to keratin (AE1 and NCL-5D3), vimentin, neurofilament, muscle specific actin, S-100 protein, neuron specific enolase (NSE), glial fibrillary acidic protein (GFAP) and others showed strong positive results. In a routine celloidin embedded human temporal bone section that was immunostained with monoclonal anti-keratin antibodies NCL-5D3  
35 and AE1 following treatment with aldehyde releasing reagent, all epithelial cells lining the cochlear duct were distinctly labeled by the anti-keratin antibodies. Keratin localization within the organ of Corti was also discernible. The keratin

immunoreactivity showed so distinctively that all epithelial cells were precisely demonstrated by the immunohistochemical staining.

Other celloidin embedded temporal bone sections immunostained with monoclonal antibodies after treatment with aldehyde releasing reagent solution also showed strong positive results. Antibody against neuron specific enolase (NSE) was localized in spiral ganglion neurons and neurofibers. Anti-NSE immunostaining was localized within inner hair cells in the organ of Corti but was absent in outer hair cells except at the bottom of the outer hair cells where it occurred possibly in synapses and terminal nerve branches. Glial fibrillary acidic protein (GFAP) was localized along the glial-Schwann junction to the brain side only. Desmin was localized in tensor tympanic muscle by anti-desmin antibody. This labelling with anti-desmin antibody was not possible using micro-dissection methods (Bauwens *et al.*, *Acta Otolaryngol.* (1990) Suppl. 470:34). The skin and appendages of dermis in the external auditory canal were also stained positively by some antibodies such as keratin and actin. Tubulin was widely localized in most epithelial and mesenchymal cells of whole temporal bone.

**Table 2. Immunohistochemical Staining on Routine Processed, Celloidin-Embedded Human Temporal Bone Sections**

	Monoclonal Antibody	Results
	Keratin:	
	AE1	+++
	NCL-5D3	+++
	Vimentin	+++
25	NF	+++
	GFAP	+++
	Desmin	++
	Myoglobin	++
	a-Tubulin	++
30	b-Tubulin	+
	Muscle Specific Actin	+++
	Chromogranin	-
	a-Actinin	-
	EMA	-
35	NSE	++
	S-100	+++

Immunoreactivity was scored on a scale of - to +++, - being non-reactive and +++ being highly reactive.

**EXAMPLE 2: AN OXIDIZING AGENT AS AN ALDEHYDE  
RELEASING REAGENT**

1. Preparation of Solution

5 Sodium hypochlorite (NaClO) in distilled water solution provides one formulation of the basic reagent used in these experiments. NaClO, 0.01-0.005 grams, was added to 100ml of distilled water in a brown colored bottle. The solution was mixed by shaking and storing at room temperature.

10 2. Treatment of Tissue Sections

The deparaffinized tissue slides were washed in distilled water for 10 minutes. The slides were immersed in an aldehyde releasing reagent solution, i.e., dilute sodium perchlorate (aqueous) solution for 10 minutes.

15 3. Immunostaining Procedures and Results

Treatment was followed by a three-step immunostaining technique. Briefly, slides were incubated with primary antibody overnight at room temperature followed by a 20-30 minute incubation with a link (BioGenex Super Sensitive biotinylated anti-mouse immunoglobulin or Vector biotinylated anti-mouse immunoglobulin). A label (BioGenex Super Sensitive Alkaline phosphatase conjugated streptavidin or peroxidase conjugated streptavidin and Vector ABC) was added for 20-30 minutes. Between incubations, slides were rinsed in three changes of PBS for 15 minutes. Either fast red or DAB chromogen was used as a substrate. The immunostaining results were controlled by light microscopy.

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25 Positive results were seen.



EXAMPLE 3: AN ORGANIC ACID/BASE PAIR AS AN ALDEHYDE  
RELEASING REAGENT

The deparaffinized tissue slides were washed in distilled water for 10 minutes.  
5 The slides were loaded with a 5-10% aqueous solution of a citric acid. After waiting for 10-30 minutes, the slides were loaded with a 10% methanolic/aqueous solution of sodium hydroxide (prepared as set forth in Example 1) for 10-30 minutes. Immunostaining proceeded as set forth in Example 2 and positive results were seen.

10 The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the appended claims.

**WE CLAIM:**

1. A method for restoring immunoreactivity of a tissue fixed with an aldehyde fixing agent and embedded in an embedding medium, said method comprising the steps of:
  - a) contacting said tissue with a solvent for said embedding medium and an aldehyde releasing reagent;
  - b) removing aldehyde released by said aldehyde releasing reagent from contact with said tissue by reacting said aldehyde in a substantially irreversible manner to form a non-aldehyde derivative; and
  - c) removing or neutralizing excess aldehyde releasing reagent from said tissue.
2. The method of Claim 1, wherein said solvent is selected from the group consisting of methanol, ethanol, propanol, butanol, ethylene glycol, and glycerol.
3. The method of Claim 1, wherein said aldehyde releasing reagent is selected from the group consisting of nucleophilic bases, oxidizing agents and organic acid/base pairs.
4. The method of Claim 1, wherein said aldehyde releasing reagent is an oxidizing agent.
5. The method of Claim 1, wherein said aldehyde releasing reagent is a nucleophilic base.
6. The method of Claim 1, additionally comprising in step a) contacting said tissue with a chaotropic agent.
7. The method of Claims 5 and 6, wherein said tissue comprises decalcified tissue.

8. A kit for use in immunostaining of a tissue fixed with a formaldehyde fixative agent and embedded in an embedding medium, said kit comprising:
- a) a first container containing an aldehyde releasing reagent solution comprising a solvent and an aldehyde releasing reagent selected from the group consisting of nucleophilic bases, oxidizing agents and organic acid/base pairs, and
  - b) a second container containing (1) an immunostaining reagent or (2) a wash solution for removing excess aldehyde releasing reagent.
9. The kit of Claim 8 wherein said aldehyde releasing reagent solution comprises a nucleophilic base, said embedding medium comprises celloidin, said immunostaining reagent comprises an antibody and said tissue is decalcified.
10. Use of an aldehyde releasing reagent selected from the group consisting of nucleophilic bases, oxidizing agents and organic acid/base pairs in a method for restoring immunoreactivity of aldehyde fixed tissue.

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/07550

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC(5) : G01N 1/28, 33/48, 33/53

US CL : 436/63, 175, 519

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 436/63, 175, 519

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, Dialog files 5 and 351.

Search terms: antigenicity, immunoreactivity, restoring, retrieval.

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	The Journal of Histochemistry and Cytochemistry, Volume 39, Number 6, issued 1991, Shi et al., "Antigen retrieval in formalin-fixed, paraffin-embedded tissues: An enhancement method for immunohistochemical staining based on microwave oven heating of tissue sections", pages 741-748. See entire article.	1-10
Y	Carey et al., Advanced Organic Chemistry, second edition, published 1983 by Plenum Press (New York). See pages 58-62.	1-10



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	* T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A* document defining the general state of the art which is not considered to be part of particular relevance	* X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* E* earlier document published on or after the international filing date	* Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* L* document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	* A*	document member of the same patent family
* O* document referring to an oral disclosure, use, exhibition or other means		
* P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

22 September 1993

Date of mailing of the international search report

13 OCT 1993

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## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/07550

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	The Journal of Biological Chemistry, Volume 174, issued 1948, Fraenkel-Conrat et al., "Reaction of formaldehyde with proteins. VI. Cross-linking of amino groups with phenol, imidazole, or indole groups", pages 827-843, see especially page 827.	1-10